

# **Plant Archives**

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# EFFECT OF PB STRESS ON RELATIVE WATER CONTENT, PHOTOSYNTHETIC PIGMENTS, PB UPTAKE AND NUTRIENTS (CA, NA AND K) BALANCE IN BROAD BEAN (*VICIA FABA* L.) PLANT

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ABSTRACT
 Lead is widely used in everyday life, it has important physicochemical properties, and it is a very useful chemical element, abundant but dangerous. In plants, it has no physiological role and is a toxic element. To study the effect of this pollutant physiologically and chemically in bean (*Vicia faba* L.) plants, different doses of lead in the form of [Pb(NO<sub>3</sub>)<sub>2</sub>] (0, 50, 100, 200, 500, 1000, 2000 ppm) were added to the culture substrate consisting of sand and compost. The results of this study showed a reduction in relative water content (RWC) in dry weight and chlorophyll pigment content. The results also showed a significant reduction in sodium (Na) in the roots and stems. The presence of lead caused significant physiological and chemical disturbances.
 *Keywords* : *Vicia faba*, lead, chlorophyll, calcium, potassium, sodium.

### Introduction

Lead (Pb) is a metal that has been used by humans for several thousand years (Gupta *et al.*, 2020), its consumption continues to decrease and its main use is in batteries (Külahcı, 2020). Lead is present in the soil, but also in water, in the air, and in living organisms, it is the most common and toxic element affecting living organisms in the environment (Shahid *et al.*, 2011). In fact, after arsenic, lead is considered the most hazardous pollutant (ATSDR, 2020). Due to its important physicochemical properties, lead is widely used in daily life, it is a very important, abundant but dangerous chemical element.

Lead is not very mobile in natural environments and tends to accumulate in the surface horizons of soils (Baize, 2010). Lead has no physiological role and remains toxic even at low doses (Beak *et al.*, 2006). Lead is a non-biodegradable element, persists in the environment and can therefore be transmitted to plants and humans via soil, air, and water (Natasha *et al.*, 2020).

The effects of lead on plants can vary depending on the duration of exposure, the intensity of the stress, the stage of development of the plant, the organs considered, and obviously the species and even the cultivar (Pourrut *et al.*, 2011). In high doses, lead leads to the total inhibition of germination, reduces the development of the seedling and the rootlets (Islam*et al.*, 2007; Dey *et al.*, 2007; Gichner *et al.*, 2008; Gopal and Rizvi, 2008), reduces the protein pool (Jana and Choudhuri, 1982; Mohan and Hosetti, 1997; Saxena *et al.*, 2003; Mishra *et al.*, 2006; Garcia *et al.*, 2006; Piotrowska *et al.*, 2009), and alters the qualitative composition of proteins (Beltagi, 2005). Lead can affect enzyme activity and

causes dysfunction of a very large number of enzymes involved in various metabolic processes (Mitra *et al.*, 2020). It leads to strong inhibition of photosynthesis, photosynthetic yield, and limitation of CO<sub>2</sub> assimilation rate (Xiong *et al.*, 2006; Hu *et al.*, 2007; Liu *et al.*, 2008; Piotrowska *et al.*, 2009; Singh *et al.*, 2010; Cenkci *et al.*, 2010).

Lead also affects plant mineral nutrition, disrupting nutrient uptake and transport (Chatterjee *et al.*, 2004; Sharma and Dubey 2005; Gopal and Rizvi 2008). Genetically the toxic effects of lead on plants result in DNA damage (Pourrut *et al.*, 2011; Shahid *et al.*, 2011; Kumar *et al.*, 2017) and chromosomal abnormalities (Rodriguez *et al.*, 2013).

The choice of the plant is based on its interest from an agronomic and eco-toxicological point of view. Thus, the plant used the bean, an annual legume botanically known as *Vicia faba* L. (Hanelt and Mettin, 1989). The bean is used for food and feed (Singh and Bhatt, 2012). The global production of dried faba bean seeds for the year 2020 has been estimated at 5.67 million tons, with China (30.4%), Ethiopia (18.89%), and the United Kingdom (9.77%) being the major producers (TRIDGE, 2022). In addition to its agronomic interest, the bean is used in studies of the response to different stresses (Marcato-Romain *et al.*, 2009).

This work consists to evaluate the impact of the presence of lead on the behavior and development of the bean. This is established by studying the application of different doses of lead in the form of  $Pb[(NO_3)]_2$ , in the culture substrate on the plants of the bean (*Vicia faba* L.). The reactions of the plants subjected to this stress are evaluated by physiological and chemical parameters.

Effect of Pb stress on relative water content, photosynthetic pigments, Pb uptake and nutrients (Ca, Na and K) balance in broad bean (*Vicia faba* L.) plant

### **Materials and Methods**

#### Plant material and experimental conditions

The plants used in this experiment are faba-bean plants *Vicia faba* L. major of the cultivar (super aguadulce).

The disinfected seeds were germinated for one week and then transplanted in a culture substrate composed of compost and sand 1V/3V. The plants were kept in the greenhouse until the end of the experiment.

Lead was added to the substrate after 5 days of transplanting the bean seedlings as lead nitrate  $Pb[(NO_3)]_2$ . The treatment levels selected were D1 = 50 ppm; D2 =100 ppm; D3 = 200 ppm; D4 = 500 ppm; D5 = 1000 ppm; D6 = 2000 ppm and the control D0 = 0 ppm.

### **Plant analysis**

#### **Physiological parameters**

30 days after the application of stress the aerial (ADW) and root dry weight (RDW) of the plants is established after removal of the plants and each part is dried at 120°C for 24 hours. The relative water content (RWC) is determined by the equation of Barrs and Weatheley (1968):

 $RWC(\%) = [(FW - DW)/(TW - DW)] \times 100$ 

TW: turgescent weight (g); FW: fresh weight (g); DW: dry weight (g)

#### **Chlorophyll pigments**

Chlorophyll pigment concentrations (chlorophyll a, chlorophyll b, and carotenoids) are determined by spectrometry (Wang *et al.*, 2010) and contents are expressed in mg/g fresh matter using the following formulas:

Chlr.a = 
$$12.25 \times A663 - 2.79 \times A645$$
(chlorophyll a)  
Chlr.b =  $21.50 \times A645 - 5.10 \times A663$  (chlorophyll b)  
CRTN =  $(1000 \times A470 - 1.82 \times Chlr.a - 85.02 \times Chlr.b)/$   
198 (carotenoids).

## **Mineral elements**

The concentrations of mineral elements calcium (Ca), potassium (K), and sodium (Na) are determined by spectrometry according to the method described by Martin-Prevel *et al.* (1984) after the destruction of the organic matter by calcination and reading with a flame spectrophotometer.

The determination of lead content is performed by atomic absorption spectrometry (AAS) after wet digestion with nitric acid and perchloric acid according to the method described by Vogel-Milkus *et al.* (2005).

## Statistical analysis

The results are presented as means  $\pm$  standard deviation (Mean $\pm$ SD). All the data obtained were statistically studied using a one-way (ANOVA), and a simple linear regression with the presentation of R<sup>2</sup>. Means are compared according to Dunnett's test at p < 0.05.

#### **Results and Discussion**

**Table 1 :** Results of the effect of lead on the tested physiological parameters

		Control	D1	D2	D3	D4	D5	D6	P value
chlorophyll a	Mean	1.20	1.00 <sup>ns</sup>	0.69**	0.48****	$0.47^{****}$	0.38****	0.33****	p<0.0001
(mg/g)	± SD	0.13	0.26	0.17	0.12	0.14	0.13	0.11	
chlorophyll b	Mean	0.83	$0.59^{**}$	$0.52^{***}$	0.31****	$0.28^{****}$	$0.17^{****}$	$0.17^{****}$	p<0.0001
(mg/g)	± SD	0.12	0.14	0.09	0.04	0.09	0.02	0.08	
carotenoids	Mean	0.56	0.53 <sup>ns</sup>	0.51 <sup>ns</sup>	0.52 <sup>ns</sup>	0.47 <sup>ns</sup>	0.46 <sup>ns</sup>	$0.42^{*}$	p<0.0654
(mg/g)	± SD	0.08	0.04	0.01	0.08	0.09	0.03	0.06	
ADW (g)	Mean	11.30	7.66**	6.94***	6.52***	6.11****	5.39****	3.75****	p<0.0001
	± SD	1.62	0.82	1.60	1.44	1.84	0.47	0.60	
RDW (g)	Mean	4.13	3.41 <sup>ns</sup>	3.08 <sup>ns</sup>	2.63**	1.68****	1.45****	0.96****	p<0.0001
	± SD	1.01	0.53	0.29	0.49	0.30	0.48	0.19	
RWC (%)	Mean	85.56	77.56 <sup>ns</sup>	75.71 <sup>ns</sup>	72.30*	71.37**	73.48*	68.64**	p<0.0103
	± SD	6.17	7.15	4.50	3.72	4.18	5.13	7.56	
Pb (mg/g)	Mean	0.00	0.05****	0.08****	0.11****	0.23****	0.36****	0.71****	p<0.0001
	± SD	0.00	0.00	0.00	0.00	0.00	0.00	0.02	
( <sup>ns</sup> ) not significa	nt; ( <sup>*,**,*</sup>	***,****) Signi	ficant (Dun	nett test)					

(p) p value not significant; (p) p value significant (one-way ANOVA test)





**Fig. 1:** Evolution of K, Na and Ca content in the aerial (A) and root (R) parts of the bean. (<sup>ns</sup>) not significant; (\*\*\*\*\*\*\*\*\*); Significant (Dunnett test) (p) p-value not significant; (**p**) p-value significant (one-way ANOVA test)

## Dry weight

The results obtained show that the increase of the lead dose applied in the substrate is accompanied by a decrease in the dry weight (Table 1), these decreases affect both parts of the plant (root and aerial). In the roots, the weight reductions are more than 17% at 50 ppm dose (D1) and reach more than 76% at D6 dose (2000 ppm). The same trend is observed for the dry weight of the aerial part where the reductions are more than 32% at D1 dose (50 ppm), and more than 66% at D6 dose (2000 ppm) in comparison with the controls (Table 1).

These results are confirmed by those found by Islam *et al.* (2008); Kopittke *et al.* (2007); Gupta *et al.* (2010) who reported a very significant decrease in the growth of the underground and aerial parts of plants even at low doses. The obtained results are in agreement with those found by Mroczek-Zdyrska *et al.* (2017) with very significant (p<0.05) reductions in fresh and dry weight of roots, in *Vicia faba* plants treated with 50  $\mu$ M Pb. Islam *et al.* (2008) reported a reduction in fresh weight up to more than 50%, compared to the control, with significant inhibition of plant development.

Root growth is more influenced by Pb toxicity than aerial parts (Zhou *et al.*, 2016). The inhibition of plant growth by decreasing biomass could be from the disruption of cell division (Rucińska *et al.*, 2004; Kozhevnikova *et al.*, 2009).

#### **Relative Water Content (RWC)**

The observation of the RWC (Tab.1) shows that increasing the Pb dose causes a decrease in RWC in the bean plants. These reductions are significant from the 200 ppm dose (D3) with more than 15%, and more than 19% reduction at the 2000 ppm dose (D6). The analysis of variance (Tab.1) records a significant effect (p<0.0103) of the Pb dose added to the substrate on RWC. The dependence between lead stress and RWC is highly significant (Fig. 2) with R<sup>2</sup>= 0.2351.

Disruptions in the water status of plants exposed to different concentrations of lead are reported in various crops (Sharma and Dubey, 2005). On *Helianthus annuus* L. plants (Kastori *et al.*, 2008), on wheat (Alsokari and Aldesuquy, 2011), and on faba bean (Pourrut, 2008). The reduction in RWC is attributed to reduced leaf area in *Glycine max* L.

plants (Elzbieta and Miroslawa, 2005), and attributed to the limitation in transpiration for to reduce Pb translocation from roots to aerial parts (Wang *et al.*, 2013).

## Photosynthetic pigment content

The results obtained (Tab.1) show that the chlorophyll a content strongly decreases with the increase of the lead dose added in the substrate. The regression of the chlorophyll a content is highly significant from the 100 ppm dose (D2) where it is more than 42%, this regression reaches more than 72% at the 2000 ppm dose (Table 1). The results obtained (Table 1) show that the fluctuations of chlorophyll a are highly dependent on the variation of the lead dose (p <0.0001<sup>\*\*\*\*\*</sup>). The observations of the results (Table 1) also show that the increase of the lead dose in the culture substrate causes a very important regression of the chlorophyll b content, these regressions are very significant starting from the 50 ppm dose (D1) where we recorded a decrease compared to the control of about 28.85% and reach at the 2000 ppm dose (D6) more than 79% (Table 1). The fluctuations in chlorophyll b content (Table 1) are dependent on the variation of the lead dose factor (p<0.0001\*\*\*\*). For Carotenoids, the results recorded (Table 1), shows a slight decline with the increase of the lead dose added in the substrate, this decrease remains insignificant until the 2000 ppm dose (D6) with a reduction of about 25.14% compared to the values recorded for the controls (Table 1). The variance of this parameter as a function of the presence of lead in the medium (Table 1) remains non-significant  $(p < 0.0654^{ns}).$ 

The inhibition of photosynthesis and reduction of photosynthetic yield in the presence of lead stress observed is confirmed by several authors (Xiong et al., 2006; Hu et al., 2007; Liu et al., 2008; Piotrowska et al., 2009; Singh et al., 2010; Cenkci et al., 2010; Azooz et al., 2011). The decreases in chlorophyll pigment observed in bean plants in this study have been reported by several works performed on the same plant (Hedaya, 2008; Pourrut, 2008; Wang et al., 2010), or on other species (Gajic'et al., 2009, Ashraf et al., 2017). The decrease in chlorophyll levels was as high as +77% (at 40 mg/L after 30 days of treatment) compared to the control in Lemna polyrrhiza L. (John et al., 2008). Ashraf et al, (2017) showed that Pb stress significantly (p<0.05) inhibits photosynthetic pigments (Chlorophyll a , b, and carotenoids) in rice plants (+87% reduction at 1200 ppm dose). In Triticum aestivum, the presence of lead at different doses (1-4 mM) in the culture medium caused very significant reductions in photosynthetic pigments (Yang et al., 2010).

# **Chemical parameters**

The recorded results of potassium content in aerial parts (Fig. 1) show a tendency to decrease slightly at dose D1 (1.89%), at dose D2 (3.61%), at dose D3 (5.92%), and at dose D5 (4.9%). The results also show an increase in potassium content in the aerial parts at dose D4 (0.93%) and at dose D6 (1.64%). For potassium in the roots, the results show a decrease with the increase in lead in the substrate (Fig. 1), this decrease is very significant from dose D2 (100ppm) with a reduction of 38% and it reaches over 56% at dose D6 (2000 ppm).

The sodium content in both parts of the plant decreases with the dose of Pb in the substrate increases. In the aerial parts, this decrease is only significant from D2 (100 ppm) 5.9%, and it reaches 19.83% for D6 (2000 ppm) (Fig. 1). In the root parts, the decrease in Na content is more important than that recorded for the aerial parts, we record (Fig. 1) for the dose D1 (50 ppm) a decrease is about 10.21% compared to the control, 26.33% for the dose D3 and more than 32% for the dose D6 (2000 ppm) (Fig.1).

The recorded results of calcium content (Fig. 1) show decreases of more than 6% for D2 and D3, more than 8% for D4 and D6, and decreases of more than 10% for D1 (50 ppm), but these decreases are non-significant p<0.3131ns (Fig. 1). Calcium in roots also decreases with increasing Pb dose, the observed decrease is only significant from dose D4 (Fig. 1) with more than 22% reduction, more than 27% at dose D5 and more than 39% at dose D6 (Fig. 1).

Pb stress disrupts the relationship between the plant and nutrients and alters the internal ratios of plant mineral elements (Gopal and Rizvi, 2008), and reduces the mineral nutrition of plants (Dotaniya *et al.*, 2020). The passage of lead through the plant membranes affects the absorption mechanism in the roots and blocks the distribution of nutrients to different parts of the plant (Sharma and Dubey, 2005).

The results presented in this work (Fig. 2) show that Pb stress induces a significant correlation with sodium in the roots ( $R^2$ =0.4315) and in the aerial part ( $R^2$ =0.6563), Pb stress also induces a significant correlation for Ca in the roots ( $R^2$ =0.6628) and for potassium in the same parts ( $R^2$ =0.4787), the correlation in the aerial parts for these two elements is not significant.

Experiments performed on wheat plants treated with different concentrations (0, 1.5, 3 and 15 mM) of Pb showed a decrease in the concentration of some macro and micronutrients (Lamhamdi *et al.*, 2013). The results obtained by Singh *et al.* (2015) showed reductions in the concentrations of calcium, sodium, and potassium in roots and aerial parts of maize plants exposed to Pb stress. Also, the reduction of potassium concentration was observed in root parts of faba bean plants exposed to mixed lead and salt stress (Azzouz and Bouziani, 2022). It was also found that Pb treatment resulted in decreased concentrations of zinc, iron, manganese, copper, calcium, phosphorus, and magnesium in *Oryza sativa* (Chatterjee *et al.*, 2004), *Medicago sativa* (Lopez *et al.*, 2007), and *Raphanus sativus* (Gopal and Rizvi, 2008).

### Lead accumulation

The observation of the results of the accumulation of lead in plants (Table 1) shows that the increase in the Pb doses applied in the substrate is accompanied by a significant increase of the lead content in the treated plants, this accumulation reaches its maximum at the dose D6 (2000 ppm) and registers more than 0.7 mg/g. The analysis of variance (Table 1) shows that the variations of the lead dose factor exert a very highly significant effect (p<0.0001) on the accumulation of lead in the plants,

The results of this work are in agreement with those obtained by Hedaya (2008) which show that the lead level in the plant increases with the concentration of lead in the medium. The work of Nadgórska-Socha *et al.* (2013) performed on faba bean showed a highly significant correlation between the presence of lead in the soil and its content in the plant organs, r=0.99 in the stem, and r=0.97 in

accumulation in the leaves of faba bean is also reported by Wang *et al.* (2007) with significant correlation r=0.97.



Fig. 2: Simple linear regression of the effect of Pb doses (ppm) on the physiological and chemical parameters tested, (R<sup>2</sup>) R squared.

# Conclusion

Under lead stress conditions, the present work showed the following:

- A significant decrease in relative water content (RWC), dry weight, and chlorophyll pigments,
- A significant reduction of Na content in the roots and aerial part and of K and Ca in the roots.
- The root parts in direct contact with Pb are the most affected by this stress.

### References

- Alsokari, S.S. and Aldesuquy, H.S. (2011). Synergistic effect of polyamines and waste water on leaf turgidity, heavy metals accumulation in relation to grain yield. *J Appl Sci Res.*, 7: 376-384.
- Ashraf, U.; Kanu, A.S.; Deng, Q.; Mo, Z.; Pan, S.; Tian, H. and Tang, X. (2017). Lead (Pb) toxicity; physiobiochemical mechanisms, grain yield, quality, and Pb distribution proportions in scented rice. *Front Plant Sci.*, 8: 259.
- ATSDR (2020). Agency for toxic substances and disease registry. Toxicological profile for lead. U.S. Department of Health and Human Services. . https://www.atsdr.cdc.gov/toxprofiles/tp13.pdf
- Azouz, M.M.; Youssef, M.M. and Al-Omair, M.A. (2011). Comparative evaluation of zinc and lead and their synergistic effects on growth and some physiological responses of Hassawi Okra (*Hibiscus esculentus*) seedlings. *American journal of plant physiogy*. 6 (6): 269-282.
- Azzouz, F. and Bouziani, E.H. (2022). Impact of salt and metallic stress on the sodium and potassium uptake by the *Vicia faba* L. plants. *Plant Archives*, 22(1): 283-287.
- Baize, D. (2010). Teneurs totales en plomb en fonction de la profondeur dans les sols «naturels», Archeo Sciences, 34 :127-135.
- Barrs, C.; Weatheley, PE. (1968). A re-examination of the relative turgidity technique for estimating water deficit in leaves. *Australian Journal of Biological Sciences*, 15: 413-428.
- Beak, K.H.; Chang, J.Y.; Chang, Y.Y.; Bae, B.H.; Kim, J. and Lee, I.S. (2006). Phyto-remediation of soil contaminated with cadmium and/or 2,4,6-Trinitrotoluene. *Journal of Environmental Biology*, 27: 311-316.
- Beltagi, M.S. (2005). Phytotoxicity of lead (Pb) to SDS-PAGE Protein Profile in Root Nodules of Faba Bean (Vicia faba L.) plants. Pakistan Journal of Biological Sciences, 8(5): 687-690.
- Bouziani, E.H.; Reguieg Yssaad, H.A. and Benouis, S. (2020). Biochemical responses of two varieties of fababeans (sidi aïch and super aguadulce) to lead stress. Plant Archives Vol. 20, Supplement 2, 2020 pp. 1278-1283.
- Cecchi, M.; Dumat, C.; Alric, A.; Felix-Faure, B.; Pradere, P. and Guiresse, M. (2008). Multi-metal contamination of a calcic cambisol by fallout from a lead-recycling plant. *Geoderma*, 144: 287-298. doi:10.1016/j.geoderma.2007.11.023.

doi:10.1016/j.geoderma.2007.11.025.

Cenkci, S.; Cigerci, I.H.; Yildiz, M.; Özay, C.; Bozdag, A. and Terzi, H. (2010). Pb contamination reduces

chlorophyll biosynthesis and genomic template stability in *Brassica rapa* L. *Environ Exp Bot*. 67(3): 467-473.

- Chatterjee, C.; Dube, B.K.; Sinha, P. and Srivastava, P. (2004). Detrimental effects of Pb phytotoxicity on growth, yield, and metabolism of rice. *Comm Soil Sci Plant An.* 35(1-2): 255-265.
- Dey, S.K.; Dey, J.; Patra, S. and Pothal, D. (2007). Changes in the antioxidative enzyme activities and lipid peroxidation in wheat seedlings exposed to cadmium and lead stress. *Brazilian Journal of Plant Physiology*, 19: 53-60.
- Dotaniya, M.L.; Dotaniya, C.K.; Solanki, P.; Meena, V.D. and Doutaniya, R.K. (2020). Lead Contamination and Its Dynamics in Soil-Plant System. In: Gupta D.; Chatterjee S.; Walther C. (eds) Lead in Plants and the Environment. Radionuclides and Heavy Metals in the Environment. Springer, Cham. https://doi.org/10.1007/978-3-030-21638-2\_5.
- Elzbieta, W. and Miroslawa, C. (2005). Pb-induced histological and ultrastructural changes in the leaves of soybean (*Glucine max* L. Merr.). *Soil Sci Plant Nutr.* 51(2):203-212.
- Gajić, G.; Mitrović, M.; Pavlović, P.; Stevanović, B.; Djurdjević, L. and Kostić, O. (2009). An aasessment of the tolerance of *Ligustrum ovalifolium* Hassk. to trafficgenrated Pb using physiological and biochemical markers. *Ecotoxicology and Environnemental Safty*, 72 : 1090-1101.
- Garcia, J.S.; Gratao, P.L.; Azevedo, R.A. and Arruda, M.A.Z. (2006). Metal Contamination Effects on Sunflower (*Helianthus annuus* L.) Growth and Protein Expression in Leaves During Development. *Journal of Agriculture and Food Chemestry*, 54(22): 8623-8630.
- Gichner, T.; Znidar, I. and Száková, J. (2008). Evaluation of DNA damage and mutagenicity induced by Pb in tobacco plants. *Mutat Res/Genet Toxicol Environ Mutagen*, 652(2): 186-190.
- Gopal, R. and Rizvi, A.H. (2008). Excess Pb alters growth, metabolism and translocation of certain nutrients in radish. *Chemosphere*, 70(9): 1539-1544.
- Gupta, D.; Huang, H.; Yang, X.; Razafindrabe, B. and Inouhe, M. (2010). The detoxification of Pb in *Sedum alfredii* H. is not related to phytochelatins but the glutathione. *J Hazard Mater*, 177(1-3): 437-444.
- Gupta, D.; Chatterjee, S. and Walther, C. (2020). Lead in Plants and the Environment. 10.1007/978-3-030-21638-2.
- Hanelt, P. and Mettin, D. (1989). Biosystematics of the genus Vicia L. (Leguminosae). Annu. Rev. Ecol. Syst. 20:199-223.
- Hedaya, A.K. (2008). Lead Accumulation and its Effect on Photosynthesis and Free Amino Acids in Vicia faba Grown Hydroponically. Australian Journal of Basic and Applied Sciences, 2(3): 438-446.
- Hu, J.; Shi, G.; Xu, Q.; Wang, X.; Yuan, Q. and Du, K. (2007). Effects of Pb<sup>2+</sup> on the active oxygenscavenging enzyme activities and ultrastructure in *Potamogeton crispus* leaves. *Russian Journal of Plant Physiology*. 54(3): 414-419.
- Islam, E.; Yang, X.; Li, T.; Liu, D.; Jin, X. and Meng, F. (2007). Effect of Pb toxicity on root morphology, physiology and ultrastructure in the two ecotypes of

*Elsholtzia argyi. Journal of Hazardous Materials*, 147(3): 806-816.

- Islam, E.; Liu, D.; Li, T.; Yang, X.; Jin, X.; Mahmood, Q.; Tian, S. and Li, J. (2008). Effect of Pb toxicity on leaf growth, physiology and ultrastructure in the two ecotypes of *Elsholtzia argyi. Journal of Hazardous Materials*, 154: 914-926.
- Jana, S. and Choudhuri, M. (1982). Senescence in submerged aquatic angiosperms: effects of heavy metals. *New Phytologist*, 90(3): 477-484.
- John, R.; Ahmad, P.; Gadgil, K. and Sharma, S. (2008). Effect of cadmium and lead on growth, biochemical parameters and uptake in *Lemna polyrrhiza* L. *Plant Soil Environ*, 54 (6): 262-270.
- Kastori, R.; Petrovic, M. and Petrovic, N. (2008). Effect of excess lead, cadmium, copper, and zinc on water relations in sunflower. *J. Plant Nutr.*, 15: 2427-2439.
- Kopittke, P.M.; Asher, C.J.; Kopittke, R.A. and Menzies, N.W. (2007). Toxic effects of Pb<sup>2+</sup> on growth of cowpea (Vigna unguiculata). Environmental pollution (Barking, Essex : 1987).150(2): 280-287.
- Kozhevnikova, A.D.; Seregin, I.V.; Bystrova, E.I.; Belyaeva, A.I.; Kataeva, M.N. and Ivanov, V.B. (2009). The effects of lead, nickel, and strontium nitrates on cell division and elongation in maize roots. *Russ J Plant Physiol.*, 56: 242-250.
- Külahcı, F. (2020). Environmental Distribution and Modelling of Radioactive Lead (210): A Monte Carlo Simulation Application. In: Gupta D.; Chatterjee S.; Walther C. (eds) Lead in Plants and the Environment. Radionuclides and Heavy Metals in the Environment. Springer, Cham. https://doi.org/10.1007/978-3-030-21638-2\_2.
- Kumar, A.; Pal, L. and Agrawal, V. (2017). Glutathione and citric acid modulates lead- and arsenic induced phytotoxicity and genotoxicity responses in two cultivars of *Solanum lycopersicum* L. *Acta Physiologiae Plantarum*. 39: 151.
- Mostafa Lamhamdi, M.; El-Galiou, O.; Bakrim, A.; Novoa-Munoz, J.C.; Arias-Estevez, M.; Aarab, A. and Lafont, R. (2012). Effect of lead stress on mineral content and growth of wheat (*Triticum aestivum*) and spinach (*Spinacia oleracea*) seedlings. Saudi Journal of Biological Sciences. 20: 29-36.
- Liu, D.; Li, T.; Jin, X.; Yang, X.; Islam, E. and Mahmood, Q. (2008). Pb induced changes in the growth and antioxidant metabolism of the Pb accumulating and non-accumulating ecotypes of *Sedum alfredii*. J Integr Plant Biol., 50(2):129-140.
- Lopez, M.L.; Peralta-Videa, J.R.; Benitez, T.; Duarte-Gardea, M. and Gardea-Torresdey, J.L. (2007). Effects of lead, EDTA, and IAA on nutrient uptake by alfalfa plants. *J Plant Nutr.* 30: 1247-1261.
- Marcato-Romain, C.; Guiresse, M.; Cecchi, M.; Cotelle, S. and Pinelli, E. (2009). New direct contact approach to evaluate soil genotoxicity using the *Vicia faba* micronucleus test. *Chemosphere*. 77: 345-350.
- Martin-Prevel, P.; Gagnard, J.; Gautier, P. and Drouineau, G. (1984). Analyse végétale dans le contrôle de l'alimentation des plantes tempérées et tropicales. Technique et Documentation-Lavoisier, p.161-179.
- Mishra, S.; Srivastava, S.; Tripathi, R.D.; Govindarajan, R.; Kuriakose, S.V. and Prasad, M.N.V. (2006). Phytochelatin synthesis and response of antioxidants

during cadmium stress in *Bacopa monnieri* L[lozenge]. *Plant Physiology and Biochemistry*. 44(1): 25-37.

- Mitra, A.; Chatterjee, S.; Voronina, A.V.; Walther, C. and Gupta, D.K. (2020). Lead Toxicity in Plants: A Review. In: Gupta D.; Chatterjee S.; Walther C. (eds) Lead in Plants and the Environment. Radionuclides and Heavy Metals in the Environment. Springer, Cham. https://doi.org/10.1007/978-3-030-21638-2\_6.
- Mohan, B.S. and Hosetti, B.B. (1997). Potential phytotoxicity of lead and cadmium to *Lemna minor* grown in sewage stabilization ponds. *Environmental Pollution*, 98(2): 233-238.
- Mroczek-Zdyrska, M.; Strubińska, J. and Hanaka, A. (2017). Selenium improves physiological parameters and alleviates oxidative stress in shoots of lead-exposed *Vicia faba* L minor plants grown under phosphorusdeficient conditions. J Plant Growth Regul., 36:186-199.
- Nadgórska-Socha, A.; Kafel, A.; Kandziora-Ciupa, M.; Gospodarek, J. and Zawisza-Raszka, A. (2013). Accumulation of heavy metals and antioxidant responses in *Vicia faba* plants grown on monometallic contaminated soil. *Environ Sci Pollut Res.* 20:1124-1134.
- Natasha, N.; Dumat, C.; Shahid, M.; Khalid, S. and Murtaza, B. (2020). Lead Pollution and Human Exposure: Forewarned is Forearmed, and the Question Now Becomes How to Respond to the Threat!.https://doi.org/10.1007/978-3-030-21638-2\_3.
- Piotrowska, A.; Bajguz, A.; Godlewska-Zylkiewicz, B.; Czerpak, R. and Kaminska, M. (2009). Jasmonic acid as modulator of Pb toxicity in aquatic plant *Wolffia arrhiza* (Lemnaceae). *Environ Exp Bot.*, 66(3):507-513.
- Pourrut, B. (2008). Implication du stress oxydatif dans la toxicité du plomb sur une plante modèle, *Vicia faba*. Thèse de doctorat de l'université de Toulouse, 284p.
- Pourrut, B.; Shahid, M.; Dumat, C.; Winterton, P. and Pinelli, E. (2011). Lead Uptake, Toxicity, and Detoxification in Plants. *Reviews of Environmental Contamination and Toxicology, Springer Verlag.* 213: 113-136.
- Rodriguez, E.; Azevedo, R.; Moreira, H.; Souto, L. and Santos, C. (2013). Pb<sup>2+</sup> exposure induced microsatellite instability in *Pisum sativum* in a locus related with glutamine metabolism. *Plant Physiol Biochem.*, 62: 19-22.
- Rucińska, R.; Sobkowiak, R. and Gwóźdź, E.A. (2004). Genotoxicity of Pb in lupin root cells as evaluated by the comet assay. *Cell Mol Biol Lett.* 9(3): 519-528.
- Saxena, A.; Saxena, D.K.; Srivastava, H.S. (2003). The Influence of Glutathione on Physiological Effects of Lead and its Accumulation in Moss Sphagnum squarrosum. Water, Air, & Soil Pollution. 143(1): 351-361.
- Shahid, M.; Pinelli, E.; Pourrut, B.; Silvestre, J. and Dumat, C. (2011). Lead-induced genotoxicity to *Vicia faba* L. roots inrelation with metal cell uptake and initial speciation. *Ecotoxicology and Environmental Safety*. 74:78-84.
- Sharma, P. and Dubey, R.S. (2005). Lead toxicity in plants. *Brazilian Journal of Plant Physiology* 17(1): 35-52.
- Singh, S.; Srivastava, P.K.; Kumar, D.; Tripathi, D.K.; Chauhan, D.K. and Prasad, S.M. (2015). Morphoanatomical and biochemical adapting strategies of maize (Zea mays L) seedlings against lead and

chromium stresses. *Biocatal Agric Biotechnol.* 4: 286-295.

- Singh, R.; Tripathi, R.D.; Dwivedi, S.; Kumar, A.; Trivedi, P.K. and Chakrabarty, D. (2010). Pb bioaccumulation potential of an aquatic macrophyte *Najas indica* are related to antioxidant system. *Bioresour Technol.* 101: 3025-3032.
- Singh, A.K. and Bhatt, B.P. (2012). Faba bean: unique germplasm explored and identified. *Hort. Flora Res. Spectrum.* 1(3):267-269.
- Tridge Exhibition 365. Plateforme d'approvisionnement mondial en produits agroalimentaires. https://www.tridge.com/fr/intelligences/broadbean/production
- Vogel-Mikus, K.; Drobne, D.; Regvar, M. (2005). Zn, Cd and Pb accumulation and arbuscular mycorrhizal colonisation of pennycress *Thlaspi praecox* Wulf. (Brassicaceae) from the vicinity of a lead mine and smelter in Slovenia. *Environmental Pollution*. 133:233-242.
- Wang, H.Q.; Lu, S.J.; Li, H. and Yao, Z.H. (2007). EDTAenhanced phytoremediation of lead contaminated soil

by Bidens maximowicziana. J. Environ. Sci. 19:1496-1499.

- Wang, J.; Li, W.; Zhang, C. and Ke, S. (2010). Physiological responses and detoxific mechanisms to Pb, Zn, Cu and Cd in young seedlings of *Paulownia forunei*. Journal of Environmental Sciences. 22(12):1916-1922.
- Wang, J.; Chen, J. and Pan, K. (2013). Effect of exogenous abscisic acid on the level of antioxidants in *Atractylodes macrocephala* Koidz under lead stress. *Environ Sci Pollut Res.* 20:1441-1449.
- Xiong, Z.; Zhao, J. and Li, M. (2006). Lead toxicity in *Brassica pekinensis* Rupr.: Effect on nitrate assimilation and growth. *Environmental Toxicology*, 21(2):147-153.
- Yang, Y.; Wei, X.; Lu, J.; You, J.; Wang, W. and Shi, R. (2010). Lead-induced phytotoxicity mechanism involved in seed germination and seedling growth of wheat (*Triticum aestivum L.*). *Ecotox Environ Safet*. 73:1982-1987.
- Zhou, C.; Huang, M.; Li, Y.; Luo, J. and Cai, L.P. (2016). Changes in subcellular distribution and antioxidant compounds involved in Pb accumulation and detoxification in *Neyraudia reynaudiana*. *Environ Sci Pollut Res.* 23:21794-21804.